ANCA Test System

RESULTS
Positive
A positive result is reported when the cytoplasm of the human granulocyte substrate displays 1+ or greater fluorescence. Differentiation between C-ANCA, P-ANCA and possible cross reactivity with GS-ANA (Granular Specific ANA) should be made. C-ANCA is defined as an uneven granular staining of the cytoplasm. P-ANCA is defined as a perinuclear / nuclear staining, and is best differentiated on an ANCA IFA slide specifically prepared for P-ANCA detection (cat no: SA007). P-ANCA at the screening dilution of 1:20 may appear dissociated on a standard ANCA slide (cat no: SA006/SA012/SA012). This reaction is caused by the inability of the ethanol fixation for C-ANCA to fix the P-ANCA antigen. C-ANCA and P-ANCA may occur together.
C-ANCA antibodies are associated with classic Wegener's granulomatosis.
P-ANCA (MPO) antibodies are associated with renal limited diseases.

Negative
A serum is considered negative for ANCA if the cytoplasm fluorescence is less than 1+. Patients should be screened on ANA HEp2 substrate to avoid confusion with PSEUDO-ANCA. PSEUDO-ANCA will stain the cytoplasm of HEp2 cells whereas true ANCA will be negative on HEp2 unless the patient possesses both ANA and ANCA antibodies.

Positive results can be confirmed on an appropriate ELISA.

TEST LIMITATIONS
1. No diagnosis should be based on a single serologic test since various host factors must be taken into consideration.

LITERATURE REFERENCES

INSTRUCTIONS FOR USE
Anti-Neutrophil Cytoplasm Antibody Test System
- Ethanol Fixed (C-ANCA) - SA060 60 Tests
- Formalin Fixed (P-ANCA) - SA070 60 Tests
Also for: Human granulocyte slides (ethanol fixed) SA006 (6 well); SA012 (12 well)
Human granulocyte slides (formalin fixed) SA007 (6 well)
C-ANCA Positive control C023N / C023N-0.5
P-ANCA Positive control C024N / C024N-0.5
ANCA Negative control C025N / C025N-0.5
FITC Anti-IgG Conjugate for ANCA J501A

INTENDED USE
The Bio-Diagnostics Anti-Neutrophil Cytoplasm Antibody (ANCA) Test kits are immunofluorescent antibody (IFA) tests to detect the presence of ANCA antibodies in human serum.

SUMMARY AND EXPLANATION
Autoantibodies specific for Wegener’s granulomatosis (WG) were first reported in the late 1970’s. Antibodies against the cytoplasmic components of the neutrophil granulocyte have also been detected in diseases other than WG, such as glomerulonephritis and/or systemic vasculitis, microscopic polyarteritis and idiopathic rapid progressive glomerulonephritis (1,2).

Standard IFA methods allow for the observation of several different patterns. Two patterns that have been well defined are C-ANCA (Cytoplasm) and P-ANCA (Perinuclear). The C-ANCA pattern shows an uneven granular staining of the cytoplasm. The P-ANCA has a perinuclear / nuclear staining pattern. During the 2nd International ANCA Workshop it was agreed that these two different patterns should be used to sub-classify the antibodies (7).

ANCA antibody detection by IFA methods has been a useful aid in the assessment of patient diagnosis, and to a certain extent their prognosis and response to therapy.

PRINCIPLE OF THE TEST
The primary reaction in this assay involves human antibody (patient sera) and a specific antigen (human granulocytes). If ANCA antibody is present in the patient sera it will bind to form an antigen/antibody complex. This occurs during the incubation period whilst the serum covers the antigen surface. A rinsing period is followed by a secondary reaction. The reagent used in the secondary reaction is a fluorescein labelled anti-human globulin conjugate. The antigen surface is then thoroughly rinsed free of unbound conjugate and viewed under an appropriate fluorescent microscope to visually identify various morphological patterns of nuclear fluorescence.
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WARNINGS AND PRECAUTIONS
1. All human components have been tested by radioimmunoassay for (HBsAg) and HTLVIII/LAV by an FDA approved method and found to be negative (not repeatedly reactive). However, this does not assure the absence of (HBsAg) or HTLVIII/LAV. All human components should be handled with appropriate care.
2. The conjugates and controls included in the kit contain 0.1% sodium azide or 0.05% Proclin as preservatives. Although these are at low concentrations, these reagents should be considered toxic. They should not be ingested or allowed to come into contact with either the skin or the mucous membranes. Sodium azide may also cause the formation of potentially explosive lead or copper azides in sinks.
3. Do not use components beyond their expiration date.
4. Follow the procedural instructions exactly as they appear in this insert to ensure valid results.
5. For in vitro diagnostic use only.
6. Handle slides by the edges since direct pressure on the antigen wells may damage the antigen.
7. Once the procedure has started do not allow the antigen in the wells to dry out. This may result in false negative test results, or unnecessary artefacts.

KIT CONTENTS
SA060/SA0150/SA0120 kit:
- SLIDE: ANCA (human granulocyte - ethanol fixed) substrate antigen slides (SA006/SA012)
- CONTROL: + C-ANCA Positive Control no: C023N / C023N-0.5

SA070 kit:
- SLIDE: ANCA (human granulocyte - formalin fixed) substrate antigen slides (SA007)
- CONTROL: + P-ANCA Positive Control no: C024N / C024N-0.5

In both kits:
- CONJ: FITC Conjugate with Evans Blue Counterstain: J501A. This reagent contains conjugates and antibodies to antigens of interest.
- CONTROL: ANCA Negative Control no: C025N / C025N-0.5
- B/A/D/A: PBS Buffer Pack no: R002
- MM: Mounting Medium no: R009

Note: All kit components are available separately. Please see the Bio-Diagnostics Ltd catalogue for more details.

ADDITIONAL MATERIALS REQUIRED BUT NOT PROVIDED
Test tubes and rack or microtitre system
Disposable pipettes
Distilled Water
Staining Dish and Slide Forceps
Moisture Chamber
Volumetric Flask (500 ml)
Fluorescence Microscope
Paper Towels - lint free

REAGENT PREPARATION

KEY FOR OTHER SYMBOLS
Used in this instruction leaflet and on accompanying product packaging:
- Manufacturer: Contains sufficient for <n> tests
- Temperature limitation: IVD In vitro diagnostic medical device
- RFU Ready for Use

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STORAGE AND STABILITY
The IFA Test System components (except PBS) must be stored at a temperature of +2°C to +8°C. Do not freeze the test kit. The stability of the kit is as indicated by the expiry date on the packaging under the above storage conditions. This applies to unopened and opened reagents. Phosphate Buffered Saline is stable at room temperature storage. The reconstituted Buffer does not contain preservatives and should be stored at 2-8°C. Care should be taken to avoid contamination.

SPECIMEN COLLECTION
Serological specimens should be collected under aseptic conditions. Haemolysis is avoided through prompt separation of the serum from the clot. Serum should be stored at 2-8°C if it is to be analysed within a few days. Serum may be held for 3 to 6 months by storage at -20°C or lower. Lipoemic and strongly haemolysed serum should be avoided. When specimens are shipped at ambient temperatures, addition of a preservative such as 0.01% thiomersal or 0.1% sodium azide is strongly recommended.

TEST INSTRUCTIONS
Screening: dilute test sera 1/20 (1 part patient sample to 19 parts diluent) in PBS. N.B. Although this dilution factor is suggested, each laboratory should determine their individual screening dilution.

Titration: set up doubling dilutions of serum starting at 1/20, (i.e. 1/20, 1/40, 1/80, 1/160, 1/320, etc.).
1. Once slides reach room temperature tear slide envelope at notch. Carefully remove the slide and avoid touching the antigen areas. The slide is now ready to use.
2. Place a drop of diluted serum (20 to 30µl) and controls over the antigen wells.
3. Place slide with patient’s serum and controls in a moist chamber for 30 minutes at room temperature (approximately 18-24°C).
4. Remove slide from moisture chamber and tap the slide on its side to allow the serum to run off onto a piece of paper towel. Using a wash bottle, gently rinse remaining sera from slide being careful not to aim the rinse stream directly onto the well.
5. Wash in PBS for 5 minutes. Repeat using fresh PBS.
6. Place an overlay on the lab table with absorbent side up. Remove slide from PBS and invert so that tissue side faces absorbent side of blotter. Line up the wells to blotter holes. Place slide on top of the blotter. Wipe the back of the slide with dry lint free paper towel. Apply sufficient pressure to slide while wiping to absorb buffer. Do not allow tissue to dry.
7. Deliver 1 drop (20-30µl) of conjugate per antigen well. Repeat steps 3-6.
8. Place 4-5 drops of mounting medium on slide.
9. Apply a 22 x 70 mm coverslip. Examine the slide under a fluorescent microscope.

Note: To maintain fluorescence, store mounted slide in a moisture chamber placed in a dark refrigerator.

QUALITY CONTROL
1. Positive and negative serum controls must be included in each day’s testing to confirm reproducibility, sensitivity and specificity of the test procedure.
2. The negative serum control should result in little (+) or no fluorescence. If this control shows bright fluorescence, either the control, antigen, conjugate or technique may be at fault.
3. The positive serum controls should result in bright 3+ to 4+ fluorescence. If these controls show little or no fluorescence, either the control, antigen, conjugate or technique may be at fault.
4. In addition to positive and negative serum controls, a PBS control should be run to establish that the conjugate is free from non-specific staining of the antigen substrate. If the antigen shows bright fluorescence in the PBS control repeat using fresh conjugate. If the antigen still fluoresces, either the conjugate or antigen may be at fault.